

SeroELISA™ Chlamydia TRUE IgM™

Enzyme-Linked Immunosorbent Assay (ELISA) for the detection of specific IgM antibodies to Chlamydia in human serum

Instruction Manual

Test kit for 96 determinations (Catalog No. 112-01)

For In Vitro Diagnostic Use For professional use only Store at 2-8°C. Do Not Freeze

Savyon® Diagnostics Ltd.

3 Habosem St. Ashdod 7761003 **ISRAEL**

Tel: +972.8.8562920 Fax: +972.8.8523176

E-mail: support@savyondiagnostics.com

Intended Use

The SeroELISA™ Chlamydia TRUE IgM™ kit is intended for the detection of specific IgM antibodies to Chlamydia in a single human serum sample by an Enzyme-Linked Immunosorbent Assay (ELISA).

For In Vitro Diagnostic Use.

Introduction

Chlamydia is a gram-negative obligate intracellular bacteria that causes acute and chronic diseases in mammalian and avian species. The genus Chlamydia is comprised of four species: C.trachomatis, C.pneumoniae, C.psittaci and C. pecorum.

C.trachomatis is divided into 15 serovars (1). Serovars A, B, Ba and C are agents of trachoma (2), the leading cause of preventable blindness endemic in third world countries. Serovars L₁-L₃ are the agents of lymphogranuloma venereum. Serovars D-K are the common cause of sexually infection transmitted genital worldwide: cervicitis, endometritis/ salpingitis (3) in females and urethritis (4) in both males and females. Endometritis/salpingitis can lead to tubal occlusion with a higher risk of extra uterine pregnancy and infertility. Genital infection may cause an acute and persistent infection occasionally without any clinical symptoms. Generally, these infections are treatable once they are diagnosed. However without any treatment the infection may progress to a severe chronic inflammation leading to infertility, ectopic pregnancy, induced abortion or premature delivery. Moreover, infants to infected mothers may be infected during birth, leading to conjunctivitis or pneumonia (5).

C.pneumoniae is an important respiratory pathogen in humans and causes up to 10% of community-acquired pneumonia cases. It has been associated with acute respiratory diseases, pneumonia, asthma, bronchitis, pharyngitis, acute chest syndrome of sickle cell disease, coronary heart disease, and Guillain-Barre syndrome (6).

C.psittaci infects a diverse range of host species from molluscs to birds to mammals and also causes severe pneumonia.

Serodiagnostic tests, which rely on specific immunologic markers, serve as a non-invasive diagnostic tool in identification of both distal and deep infections (7). It has been found that ANTI-Chlamydia IgM antibody is of diagnostic value in pneumonia caused by *C. pneumoniae* (TWAR) and *C.psittaci* (8). The serological pattern of Chlamydia IgM in chlamydial pneumonititis is as follows: antibodies are produced in the early stages of an infection, peak after 1-2 weeks and generally decline gradually to undetectable levels within 2-3 months (11).

This pattern has been observed in 20-50% of infants born to mothers who were culture positive for Chlamydia trachomatis and/or demonstrated elevated levels of specific IgG and IgA antibodies to Chlamydia. These infants developed chlamydial pneumonitis during the first 6 months of life (7).

Since IgM is present only in acute and/or recent disease (9), the Chlamydia IgM test requires only a single serum specimen and results can be reported in terms of presence or absence of immune IgM.

High titers of immune IgG, which compete with immune IgM for the same antigenic sites, may produce false negative IgM results. Rheumatoid factor (Rf. autoimmune activity) causes false positive IgM results (10). Therefore, removal of IgG and Rf is an essential part of the IgM assay.

SeroELISATM Chlamydia test employs the L₂ serovar broadly reacting antigen of C. trachomatis. It will detect C. trachomatis, C. psittaci and C. pneumoniae (TWAR) antibodies.

Principle of the test

- Human serum to be tested is brought into contact with the antigenic material coating the microtiter wells. Specific antibody, if present in the patient serum will bind to the attached antigen, a complex is formed and all the serum components are washed away in the wash phase.
- Horseradish peroxidase (HRP) conjugated anti-human IgM (µ chain specific) is added to the wells. If an antigenantibody complex was formed in the previous step, the peroxidase conjugated antibody will bind to the antibody moiety of the complex. If no antigen-antibody complex was formed in the previous step, the conjugate is washed away in the wash phase.
- TMB-Substrate is added. A positive reaction is indicated by a blue to deep blue color which develops in the test wells following enzymatic reaction of the peroxidase moiety with peroxide and the chromogen reactant. After the enzymatic reaction is stopped by an acidic solution, the absorbance of the test wells is determined at 450nm by a spectrophotometer.
- The absorbance at 450nm is indicative of IgM anti-Chlamydia titer in patient serum specimens.

Assay Procedure

- Chlamydia Antigen Attached to Solid Phase (Ag)
- Serum positive for IgM Anti-Chlamydia (Ab₁)



AgAb₁ Complex

2. AgAb₁ Complex

HRP Conjugated
 Anti-Human IgM (Ab₂)

AgAb₁Ab₂ Complex

3. AgAb₁Ab₂ Complex - TMB-Substrate

 $oldsymbol{\Psi}$

Blue Solution

◆ Chromogen Stop Solution Yellow Solution

(Absorbance Determination at 450nm)

Warning and Precautions

 Warning: THE CHLAMYDIAL ANTIGENIC MATERIAL HAS BEEN INACTIVATED AND CONTAINS NO DETECTABLE LIVE ORGANISMS. HOWEVER, THE STRIPS SHOULD BE HANDLED AND DISPOSED OF AS WOULD ANY POTENTIALLY BIO-HAZARDOUS LABORATORY MATERIAL.

Precautions: This kit contains human sera which have been tested by FDA approved techniques, and found to be negative for HBV antigen and for HCV and HIV 1 and HIV 2 antibodies. Since no known method can offer complete assurance that products derived from human blood do not transmit infection, all human blood components supplied in this kit must be handled as potentially infectious serum or blood according to the recommendations published in the CDC/NIH manual "Biosafety in Micro Biological and Biomedical Laboratories, 1988".

- Substrate/Chromogen Solution is an irritant material to skin and mucous membranes. Avoid direct contact.
- For In-vitro Diagnostic Use.

Kit contents

 Precoated microtiter plate (96 wells per frame). Each sachet contains one microtiter plate comprising 12 removable strips in a plastic frame. Each strip is coated with Chlamydia antigens.

1 Unit

Positive Control (human serum positive for anti-Chlamydia IgM antibody). Ready to Use.

1 Vial, 2.0ml

Negative Control (human serum negative for anti-Chlamydia IgM antibody). Ready to Use.

1 Vial, 2.0ml

4. Ready To Use HRP Conjugated Anti-Human IgM (μ-chain specific).

1 Vial, 10ml

5. IgM Serum Diluent, Ready-to-Use.

2 bottle, 60ml

6. Concentrated Wash Buffer (x20).

1 bottle, 100ml

7. TMB-Substrate. Ready-to-Use.

1 vial, 16ml

8. Stop Solution (1M H₂S0₄), Ready-to-Use.

1 bottle, 16ml

9. Plate cover.

10. Instruction Manual.

1 Unit 1

Materials Required But Not Supplied

- 1. Clean test tubes for dilution of patients sera.
- Adjustable micropipettes, or multichannel pipettes (5-50, 50-200 and 200-1000µl ranges) and disposable tips.
- Disposable plastic pipettes (assorted sizes) and safety pipetting devices.
- 4. One liter volumetric flask.
- 5. One 50ml volumetric cylinder.
- 6. ELISA plate washer or wash bottle.
- 7. Paper towels or absorbent paper.
- 8. Vortex mixer.
- 9. A 37°C water bath with a lid, or a moisture chamber placed in a 37° \pm 1°C incubator.
- 10. A 4°C refrigerator.
- 11. ELISA-reader with 450nm filter.
- Distilled or double deionized water for the dilution of Concentrated Wash Buffer.

Storage and Shelf Life of Reagents

All the materials supplied should be stored at 2° to 8°C. If kept at 2° to 8°C the test reagents are stable until the expiry date indicated on the kit pack. Exposure of originally stoppered or sealed components to ambient temperature for a few hours will not cause damage to the reagents. **DO NOT FREEZE!**

When a kit is in use, the shelf life of the original material is 60 days from the day first opened. Once opened, the aluminum foil sachet containing the strips should be sealed with a tape. The dehydrating packet should not be removed.

Crystals may form in the 20x concentrated Wash Buffer during cold storage, this is perfectly normal. Redissolve the crystals by warming the buffer to 37°C before diluting. Once diluted, the solution may be stored at 2-8°C up to twenty-one days

Specimen Collection

Serum specimens should be collected aseptically and stored at 2° to 8° C with 0.05% sodium azide (NaN₃) as a preservative if they are to be tested within a few days. For longer periods, aliquots of serum specimens should be stored at -20°C.

Since turbid or hemolytic serum samples may give less reproducible results, it is highly recommended that serum samples tested be clear and non hemolytic.

Assay Procedure

Notes:

- The components of this kit have been tested as one unit.
 Do not mix components from different kit lots or other manufacturer's kits.
- b) All reagents should reach room temperature before use. Serum Diluent and Conjugate Diluent gelatinize when refrigerated. If needed, accelerate liquefaction by warming these components at 37°C for several minutes. Salt crystals may form in the Concentrated Wash Buffer when stored at 2° to 8°C. These crystals should be

- completely redissolved by warming at 37°C before dilution.
- c) Do not perform the test in the presence of reactive vapors (e.g. from acid, alkaline or aldehyde substances) or dust, since the enzymatic activity of the HRP conjugated Anti-Human IgM may be affected.
- d) Do not touch the top of the strips. Do not touch the edges of the wells with the pipette tips when dispensing reagents.
- Use disposable pipette tips. Avoid cross contamination between reagents.
- Tap vial lightly on hard surface to free liquid that might be entrapped in the cap.
- g) Avoid entrapping air bubbles in the wells.
- h) Dispense liquids slowly to avoid spraying.
- Positive Control and Negative Control sera should be run together with serum specimens each time the test is performed.
- j) One well per test should be used for a blank value each time the test is performed.
- k) All the procedure steps should be performed sequentially without interruption.

Test Procedure - Manual

Automation protocol available upon request

A) Pre- Washing of Strips

Prewashing of strips before starting the assay is advisable to decrease background noise signal.

- 1. Remove required number of strips from their aluminum foil sachets and insert them in the plate frame.
- Dilute the Concentrated Wash buffer 1:20 with distilled water. For 100ml of Wash Buffer use 5ml of Concentrated Wash Buffer with 95ml distilled water and mix well.

Wash Buffer working solution should be prepared before use and excess discarded.

- Wash the strips with the washing buffer, then discard the strip contents. Repeat this step one more time.
- 4. Dry the top of the strips and frame by gently tapping them over clean absorbent paper.

B) Incubation of Serum Samples and Controls

5. Dilute each patient serum 1:105 with the supplied Serum Diluent as follows: Add 10μl of patient serum to 200μl of Serum Diluent (1/21), and then dilute further by adding 25μl of 1/21 dilution to 100μl of Serum Diluent.

Note: The Serum Diluent contains Anti-human IgG for the removal of IgG antibodies from human serum.

Controls are provided in a ready-to use form and should not be diluted.

 Pipette 50µl of Ready to Use Positive Control, Negative Control and serum specimen (1:105 dilution) into corresponding separate wells of the test strips.
 Pipette 50µl of IoM Serum Diluent into one well for blank

Pipette 50µl of IgM Serum Diluent into one well for blank value.

- Pipetting of controls and serum specimens into the wells should not exceed 10 minutes.
- 7. Cover the strips with a plate cover and incubate for 30 minutes at 37°C in a moisture chamber.
- 8. Discard the liquid content of the wells. Wash the wells **five** times and dry as in steps A).

C) Incubation with Conjugate

- Pipette 50µl of ready to use HRP conjugated Anti-Human IgM into each well.
- 10. Cover the strips with a plate cover and incubate for 30 minutes at 37°C in a moisture chamber.
- Discard the liquid content of the wells, wash them five times and dry as in steps A).

D) Incubation with TMB Substrate

- 12. Pipette 100µl of TMB Substrate into each well.
- 13. Cover the strips with a plate cover and incubate at room temperature for 15 minutes.
- Stop the reaction by adding 100µl Stop Solution to each well.
 - Pipette the Stop Solution in the same sequence and at the same time intervals as the TMB Substrate.
- Calibrate the spectrophotometer on the blank well.
 Determine the absorbance at 450nm and record the results.

Immediate determination of the absorbance is advisable but not mandatory. Absorbance determination should not exceed 30 minutes, following stopping of chromogenic reaction.

Test validation

A test-run is valid if:

- Positive Control absorbance is ≥0.8 at 450nm
- Negative Control absorbance is ≤0.15 at 450nm
 If these conditions are not fulfilled, the test run is invalid and should be repeated.

Calculation of Cut-Off Value (COV)

The cut-off value is calculated according to the following formula:

$$COV = 0.24 \times (Pc - Nc) + Nc$$

Pc = Absorbance of Positive Control at 450nm Nc = Absorbance of Negative Control at 450nm

Interpretation of Test Results

Absorbance at 450nm	Result	Interpretation of Results	
Below COV-0.03	Negative	No detectable IgM antibodies to Chlamydia	
COV±0.03	Equivocal	Retest serum samples classified equivocal. If equivocal result is repeated testing of subsequent serum sample is recommended	
Above COV +0.03	Positive	Indication of acute and/or recent chlamydial infection	

Test Limitations

- No single serological test should be used as the only criterion for diagnosis. All clinical and laboratory data should be taken into account.
- The test is a single serovar (L₂) ELISA. L₂ contains antigenic determinants existing in serovars of Chlamydia trachomatis as well as the group antigen. Antibodies against Chlamydia psittaci, Chlamydia pneumoniae (TWAR) and Acinetobacter calcoaceticus may be detected by this ELISA.
- This test will not indicate the site of chlamydia infection(s). It is not intended to replace cell culture isolation, if available.
- Since infection with Chlamydia may not produce any immediate significant symptoms, the acute stage may be missed and there may be no detectable IgM antibodies. This does not exclude the possibility of a chlamydial infection.
- Bacterially contaminated or hyperlipaemic serum may cause erroneous results.

Performance Characteristics

SeroELISATM Chlamydia test was compared to IPAzymeTM Chlamydia TRUE- IgMTM test (Savyon Diagnostics product, Cat. No. 012-01) which is an accepted serological test for detection of IgM antibodies to Chlamydia.

The study population included patients with suspected Chlamydia infections as well as healthy individuals (n=162). The results are summarized as follows:

Comparison of SeroELISA™ with IPAzyme™

SeroELISA™ IPAzyme™	Positive	Negative	Total
Positive	76	4	80
Negative	4	78	82
Total	80	82	162

Overall agreement: $(154/162) \times 100 = 95.1\%$

Cross Reaction

Hospitalized patients, infected with *Neisseria gonorrhea*, *Staphylococcus aureus* and *Peptostreptococcus anaerobius*, who were diagnosed by commercial serology kits, were also tested with the SeroELISA Chlamydia kit. There was no significant cross-reaction detected.

Bibliography

- Yuan, Y., Zhang, Y. X., Watkins, N. G. and Caldwell, H.D. (1989). Nucleotide and Deduced Amino Acid Sequences for the Four Variable Domains of the Major Outer Membrane Proteins of the 15 Chlamydia trachomatis Serovars. Infection and Immunity. 57:1040-1049. Copyright 1989, American Society for Microbiology.
- Treharne J. D. (1985). The community epidemiology of trachoma. Rev Infect Dis. 7:760-763.
- Piura, B., Sarov, I., Sarov, B., Kleinman, D., Chaim, W. and Insler, V. (1985). Serum IgG and IgA antibodies specific for Chlamydia trachomatis in salpingitis

- patients as determined by the immunoperoxidase assay. Eur. J. Epidemiol 1: 110-116.
- Wang, S.P., Grayston, J.T., Kuo, C.C., Alexander, E.R., and Holmes, K.K. (1977). SeroDiagnosis of Chlamydia trachomatis infection with the microimmunofluorescence test. In: Nongonoccolcal urethritis and related infection, D. Hobson and K.K. Holmes (Eds), P. American Society for Microbiology, Washington DC. p. 237-248.
- Thompson III S. E., and Dretler R. H. (1982). Epidemiology and Treatment of Chlamydial Infections in Pregnant Women and Infants. Review of Infectious Diseases 4:S747
- Saikku P., Mattila, K., Nieminen, M.S., Huttunen, J.K., Leinonen, M., Ekman, M.R., Makela, P.H., and Valtonen, V. (1988). Serological evidence of an association of a novel Chlamydia, TWAR, with chronic coronary heart disease and acute myocardial infection. Lancet II: 983-986.
- Puolakkainen, M., Saikku, P., Leinonen, M., Nurminen, V., Vaananen, P. and Makela, P.H. (1984). Chlamydia Pneumonitis and its Serodiagnosis in Infants. J. Infect. Dis. 149:598-604.
- 8. **Grayson, J.G.** (1989). Chlamydia pneumoniae, Strain TWAR. Chest 95:664-669.
- Gardner, P.S. Rapid Virus Diagnosis. In Voller, A., Bartlett, A. and Bidwell, D. (Eds). Immunoassays for the 80s, pp. 353-360 MTP Press Limited 1981.
- Chantler, S. and Diment, J.A. current Status of Specific IgM Antibody Assays. In Voller, A., Bartlett, A. and Bidwell, D. (Eds). Immunoassays for the 80s, pp. 417-430 MTP Press Limited 1981.
- Numazaki, K., Chiba, S., Yamanaka, T., Moroboshi, T., Aoki, K., Nakao, T. (1985). Detection of IgM Antibodies against Chlamydia trachomatis by Enzyme Linked Fluorescence Immunoassay. J.Clin. Pathol. 38:733-739.



Obelis s.a. (European Authorized Representative Center)
Boulevard Général Wahis 53, 1030 Brussels, Belgium
Tel.: +32.2.732.59.54 Fax.: +32.2.732.60.03
e-mail: mail@obelis.net